Hygienic Laboratory

The University of Iowa

Chlamydia trachomatis/ Neisseria Gonorrhoeae

Submission Requirements

- Label each specimen transport tube with the patient name or unique identifier. Name should match that found on the Test Request Form.
- UNLABELED SPECIMENS WILL NOT BE TESTED.
- A completed Chlamydia trachomatis/Neisseria Gonorrhoeae Test Request Form must accompany each specimen.

Specimen Collection and Handling

Female Endocervical Specimen Collection

- Remove excess mucus from cervical os and surrounding mucosa using cleaning swab (white shaft swab in package with red printing). **Discard swab.**
- Insert specimen collection swab (blue shaft swab in package with green printing) into endocervical
 canal. Using the white shafted cleaning swab for specimen collection will result in specimen rejection.
- Gently rotate swab clockwise for 10-30 seconds in endocervical canal to ensure adequate sampling.
- Withdraw swab carefully; avoid any contact with vagina mucosa.
- Remove cap from swab specimen transport tube and immediately place specimen collection swab into specimen transport tube.
- Carefully break swab shaft at scoreline; use care to avoid splashing contents.
- Re-cap swab specimen transport tube tightly.
- After collection, transport and store swab in swab specimen transport tube at 2°C to 30°C until tested.

Male Urethra Swab Collection

- Patient should not have urinated for at least 1 hour prior to specimen collection.
- Insert specimen collection swab (blue shaft swab in package with green printing) 2-4 cm into urethra.
- Gently rotate swab clockwise for 2-3 seconds in urethra to ensure adequate sampling.
- Withdraw swab carefully.
- Remove cap from swab specimen transport tube and immediately place specimen collection swab into specimen transport tube.
- Carefully break swab shaft at scoreline; use care to avoid splashing contents.
- Re-cap swab specimen transport tube tightly.
- After collection, transport and store swab in swab specimen transport tube at 2°C to 30°C until tested.

Female/Male Urine Specimen Collection

- Patient should not have urinated for at least 1 hour prior to specimen collection.
- Direct patient to provide first-catch urine (approximately 20-30 mL of initial urine stream) into urine collection cup free of any preservatives. Collection of larger volumes may result in specimen dilution that may reduce test sensitivity. Female patients should not cleanse labia area prior to providing specimen.
- Remove cap from urine specimen transport tube, and transfer 2 mL of urine into tube using disposable pipette provided. Fluid level must fall between the two black indicator lines on the tube label.
- Re-cap urine specimen transport tube tightly. This is now known as the "processed urine specimen".
- Urine samples that are still in the primary collection container must be transported to the lab at 2°C to 30°C.
- Transfer urine samples into APTIMA urine specimen transport tube within 24 hours of collection. Store at 2°C to 30°C.
- One swab per transport tube. Please discard the white shaft cleaning swab.
- Transport tube caps should be tightened securely to avoid specimen rejection.
- Processed urine specimens should be assayed with the APTIMA Assay for CT and/or GC within 30 days of collection.

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- Swab specimens must be assayed with the GEN-PROBE APTIMA Assay for CT and/or GC within 60 days of collection.
- If longer storage is needed, freeze specimens at -20°C to -70°C for up to 90 days after collection.
- Wrap the specimen in absorbent material, and place into a biohazard bag.
- Use one biohazard bag per specimen.

Complete Test Request Form

- Complete a Test Request Form with the following information:
 - Name, birth date, gender, and patient ID#
 - Specimen type and date of specimen collection

Shipping Instructions

- Include completed Test Request Form in white plastic circular mailer.
- Place biohazard bag containing specimen and Test Request Form in plastic circular mailer. One to six specimens can be shipped in circular mailer.
- Use the preaddressed label provided.
- When shipping a large number of specimens, a sturdy cardboard box can be used instead of the plastic white circular mailer. This will save postage.
- Transport specimens to UHL as soon as possible after collection.
- Transport at 2°C to 27°C (room temperature).

Contact Information

- For test request forms and kits: 319/335-4466
- For additional forms, go to http://www.uhl.uiowa.edu/kitsquotesforms
- Any questions pertaining to patient test results should be directed to: Kris Eveland at kristofer-eveland@uiowa.edu or Sandy Jirsa at sandy-jirsa@uhl.uiowa.edu If it is not possible to email, call them at 319-334-4500.

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